

ANTI-MICROBIAL SENSITIVITY PATTERNS OF BACTERIAL ISOLATES FROM SURGICAL SITE INFECTIONS IN OBSTETRICS AND GYNAECOLOGY

Syeda Sitwat Fatima, Saima Gillani, Jamila Naib, Zakia Sharafat, Tayyaba Mazhar, Parveen Naveed

Department of Obstetrics & Gynaecology, Khyber Teaching Hospital, Peshawar - Pakistan

ABSTRACT

Objective: To find out the most common bacterial pathogens responsible for surgical site infections and their antimicrobial sensitivity profile.

Materials and Methods: This descriptive study was carried out in Obstetrics and Gynaecology Unit "C" Khyber Teaching Hospital, Peshawar, from June 2012 to May 2013. Samples were collected from the patients showing signs of surgical site infection, using sterile swabs and sent to laboratory for identification of micro organisms by Gram-stains and culture growth followed by in vitro antibiotic susceptibility testing.

Results: During the study period, a total of 937 patients underwent major abdominal surgeries, of them 46 patients (4.90%) developed surgical site infections. A total of 36 pathogens were detected. Staphylococcus aureus (30.4%) was the predominant organism followed by Pseudomonas aeruginosa (22.2%), Escherichia Coli (16.7%) and Enterbacter species (8.3%). Staphylococcus aureus exhibited good sensitivities to Vancomycin (100%), Amikacin (91%), Ciprofloxacin (83.3%). Low susceptibility patterns were observed to 2nd and 3rd generation Cephalosporins. Sensitivity pattern of P-aeruginosa showed 100%, 87.5%, 75% sensitivity to Imipenem, Piperacillin-Tazobactam, Amikacin and respectively. However, low susceptibility patterns were observed to Cephalosporins and amoxicillin-clavulonic acid. Escherichia Coli had good susceptibility patterns for Amikacin (100%), Imipenem (100%), Piperacillin-Tazobactam (100%), Ciprofloxacin (80%).

Conclusion: Staphylococcus aureus remains the most common pathogen involved in surgical site infections. Since a high proportion of samples had positive cultures, infection control is recommended as a strategy to minimize spread of resistant organisms.

Key Words: Surgical site, infection, pathogens, antimicrobial, sensitivity.

INTRODUCTION

Infections that occur in the wound created by an invasive surgical procedure are generally referred to as surgical site infections (SSI)¹. It is estimated that millions of surgical procedures are performed annually worldwide; SSIs continues to be a major source of morbidity after operative procedures. Despite considerable research on best practices and strides in refining surgical techniques, technological advances and environmental improvements in the operating rooms and the use of prophylactic pre-operative antibiotics, SSIs remains the second most common adverse event occurring to hospitalized patients².

SSIs are one of the most important causes of health care associated infections with incidence

according to Centers for Disease Control and Prevention (CDC) was 15.45% and according to the UK nosocomial infection surveillance was 11.32%³. In US, the reported rate of SSI is 2.6% for all operations⁴.

SSI has an adverse impact on the patient as well as on the hospital. It is responsible for prolonged hospital stay, which results in social and economic loss to the patient and the family⁵. Other clinical outcomes of SSI include poor scars that are cosmetically unacceptable, persistent pain and a significant impact on emotional well being⁶. It also adds appreciable burden on hospital resources and on health care providers⁷.

A complex interplay between host, microbial and surgical factors ultimately determines the prevention or establishment of a wound infection. The most common group of bacteria responsible for SSIs is staphylococcus aureus⁸. Other micro-organisms implicated are Escherichia-coli, Pseudomonas and Klebsiella⁹.

Address for Correspondence:

Dr. Syeda Sitwat Fatima

H. No 12 (old) Doctors Flats

Khyber Teaching Hospital,

Peshawar - Pakistan

Cell: 0333-9301667

Email: smkakakhel@hotmail.com

The wide spread use of antibiotics, together with the length of time over which they have been available has led to the emergence of resistant strains, which has significantly increased the morbidity and mortality associated with wound infections¹⁰. Routine surveillance for the hospital acquired wound infections is recommended by both the Centers of Disease Control and Prevention¹¹ and the Surgical Infection Society¹². This study is therefore, designed to determine the distribution of bacterial pathogens isolated from septic post operative wounds and their anti-microbial susceptibility patterns.

MATERIAL AND METHODS

This descriptive study was carried out in the Obstetrics and Gynaecology Unit "C" Khyber Teaching Hospital, Peshawar, from June 2012 to May 2013. All the patients who underwent major abdominal surgeries in the selected ward during the study period were included in the study. Patients were enrolled after obtaining informed consent. In all cases pre-operative, intra-operative and post-operative details were studied. Information was collected in case record form for age, date of admission, reason for admission, associated co-morbid condition/s, type of surgery: emergency or planned, the procedure performed, pre-operative antibiotic prophylaxis and the post operative day on which signs of infection were detected. All the patients who underwent major abdominal surgeries, who later on developed wound infection within 30 days, post operatively were included in the study. However, patients who had undergone major abdominal surgeries outside the Obstetrics & Gynaecology Unit "C" Khyber Teaching Hospital and were admitted for the treatment of post operative wound infection, patients with intentional reopening of wound for wound hematomas and patients undergoing diagnostic laparoscopies were excluded from the study.

Samples were collected from the wounds showing signs of infection such as erythema, tenderness, discharge and wound gaping by using a sterile swab. For collection of sample wound was washed with normal saline and then swab was collected to avoid the growth of skin commensals. The swabs were labeled with necessary details and were transported to the laboratory, where they were processed by the conventional microbiological methods and antimicrobial susceptibility was done. The data was entered into SPSS version 10 and results were obtained.

RESULTS

During the study 46 (4.90%) developed surgical site infection. Types of surgery done among the wound infected population are shown in figure 1. Prevalence of surgical site infections in relation to different types

of obstetrical and gynecological surgeries is shown in Table 1. In 33 (71.7%) of the population, wound infection appeared within 4th to 6th day post operatively. Infection appeared within 7th to 14th day and beyond 14th day post operatively in 10 (21.7%) and 3 (6.5%) cases respectively.

Among the wound infected population, 34 (74%) of patients cultures were positive for micro organism while 12 (26%) had yielded no growth. In 3 (8.8%) cases the growth was polymicrobial while in 31 (91.2%) cases only single microorganism was detected. Frequency of isolates of gram positive organisms was (41.66%) and that of gram negative organisms was (58.33%).

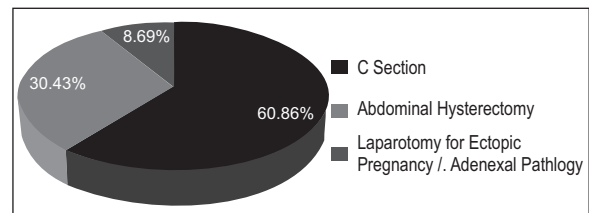


Fig. 1: Types of Surgical procedures presented with Surgical Site infection

Table 1: Prevalence of surgical site infections in relation to different types of surgical procedures

Type of surgical procedures	Frequency and per centage of surgical site infections
Emergency Caesarean sections	24 (4.35%)
Elective Caesarean sections	04 (1.82%)
Abdominal hysterectomies	14 (15.21%)
Laprotomies for ectopic pregnancy	02 (9.52%)
Laprotomies for ovarian cysts	02 (6.0%)
Myomectomies	00 (00%)
Total	46 (4.90%)

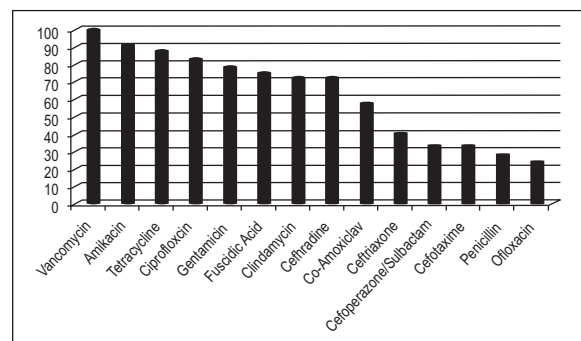


Fig. 2: Sensitivity Patterns of Staphylococcus aureus

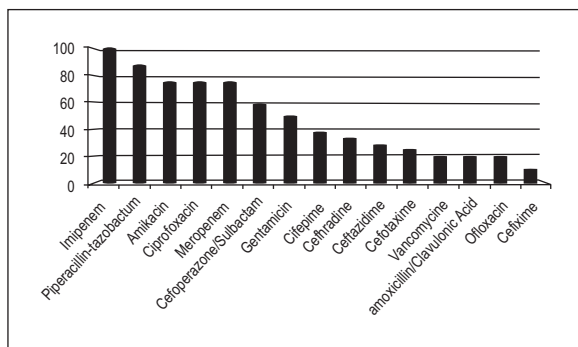


Fig 3: Sensitivity Patterns of Pseudomonas aeruginosa

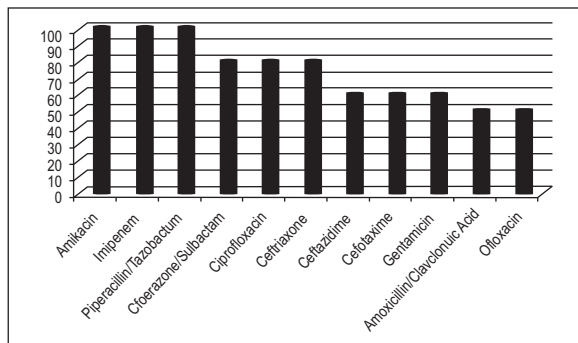


Fig. 4: Sensitivity Patterns of Escherichia coli

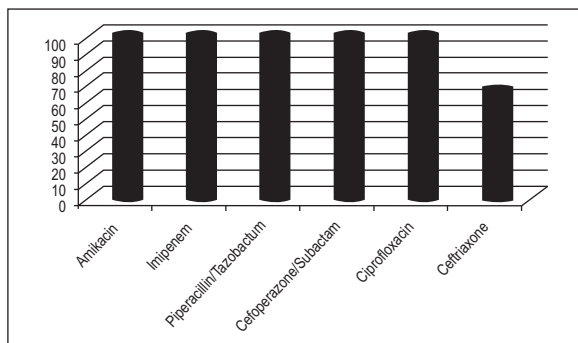


Fig. 5: Sensitivity Patterns of Enterobacter

The micro organisms isolated were Staphylococcus aureus 14 (38.9%), Pseudomonas 8 (22.2%), Escherichia coli 6 (16.7%), Enterobacter 3 (8.3%), Enterococcus 1 (2.7%), Proteus species 1 (2.7%), Klebsiella pneumoniae 1 (2.7%), Citrobacter 1 (2.7%) and Acinetobacter baumannii 1 (2.7%).

Regarding antimicrobial sensitivity patterns, Staphylococcus aureus exhibited highest sensitivities to Vancomycin (100%), Amikacin (91%), Tetracyclines (87.5%) and Ciprofloxacin (83.3%). However, very low susceptibility was seen against Penicillin (28%) and Ofloxacin (14.2%). Antimicrobial sensitivity patterns of Staphylococcus aureus is shown in figure 2. Similarly, sensitivity patterns of Pseudomonas, Escherichia coli and enterococcus shown in species are Figures 3, 4 & 5 respectively. One case of Proteus (2.7%) was detected and was sensitive to Amikacin,

Cefoperazone-sulbactam, Piperacillin-tazobactam, Gentamicin and Erythromycin. However, resistance was observed to Ceftriaxone and Ceftazidime.

Klebsiella (2.7%) was sensitive to Amikacin, Imipenem, Piperacillin-Tazobactam and Ciprofloxacin, while it was resistant to Ampicillin, Ceftriaxone, Cefuroxime, Cefixime and Gentamicin. Enterococcus (2.7%) was sensitive to Amoxicillin-clavulonic acid, Ampicillin, Gentamicin, Tetracycline and Vancomycin. However, it showed resistance to Erythromycin. Acinetobacter baumannii (n=1) was sensitive to Amikacin, Imipenem, Polymyxin-B and Meropenem. Resistance was observed to Ampicillin, Amoxicillin-clavulonic acid, Ceftriaxone, Cefixime, Ciprofloxacin, Gentamicin and Piperacillin-Tazobactam.

The analysis of Performa revealed that the prophylactic antibiotics used pre-operatively as well as post-operatively were Ceftriaxone (in 31 patients) and Amoxicillin-clavulonic acid (in 15 patients), which were available free of cost from hospital's pharmacy. It is evident that majority of the isolates are resistant to the antibiotics used for the prophylaxis and treatment of infected wounds.

DISCUSSION

In the present study incidence of surgical site infection was 4.90% among the patients who underwent major abdominal surgeries in Obstetrics and Gynecology Unit "C" Khyber Teaching Hospital, Shahida Meo⁵ from Lahore reported rate of surgical site infection of 1.92% while Nisa¹³ from Peshawar showed rate of 6.5% in her study. According to the 2004 survey report by the Centers of Disease Control and Prevention, Nosocomial Infection National Surveillance Service (NNIS) incidence of hospital acquired infection, related to surgical wounds is 0.5-13%¹⁴.

In our study surgical site infections have occurred more in elective surgeries i.e., abdominal hysterectomies (15.21%) than in emergency Caesarean-sections (4.35%). This observation may seem surprising as emergency cases have known to land up more in surgical site infection than the elective ones. However, Varsha¹⁵ has shown increased surgical site infection rate in elective gynecological surgeries as compared to emergency procedures. All of the patients undergoing abdominal hysterectomies were above 45 years of age. The presence of underlying condition such as diabetes mellitus (64.28%) and anemia, requiring blood transfusions (54.54%) were more in patients who underwent elective surgeries. Moreover, prolonged hospital stay pre-operatively in elective cases increases the risk of nosocomial infections in these patients. The above mentioned factors could have been responsible for this unexpected outcome.

Of the 46 SSIs cases 26% had yielded no growth of microorganisms, varsha¹⁵ and Rehman⁹ have also reported negative cultures in 22% and 35% cases, respectively in their studies. This may be due to the normal healing process where the bacteria have been overpowered by body's defense mechanism. Another possible explanation is anti microbial activity in patients' circulation, since all of them had been on antibiotic therapy postoperatively at time of collecting the specimen. It may be possible that the culprit microorganisms were anaerobic bacteria that were missed as cultures were incubated aerobically¹⁶.

In the present study and some other recent studies^{10,15}, predominance of gram negative organisms in surgical site infection (58.33% vs. 41.66%) is reported. However, a study done in Turkey¹⁷ showed gram positive organisms as predominant. This difference may be due to variation in common noscomial pathogens in different hospital set ups. Though, the percentage of gram negative bacilli from SSIs was high, however, *Staphylococcus aureus* was the predominant micro-organism isolated (38.88%), followed by *Pseudomonas* (22.22%) and *E-coli* (16.66%). Similar findings have been observed in the studies done previously^{18,19,20}. However, other studies have shown the predominance of *Pseudomonas*²¹, *E-Colli*¹⁵ and *Klebsiella*⁷. The high prevalence of *Staphylococcus aureus* in the present study is may be due to the fact that it forms a part of the normal skin flora and can be isolated from noses of up to 60% of the healthy individuals. It is readily transmitted from person to person, onto the hands and clothes of the health care staff, on objects and into the air²².

In present study, prevalence of Methicillin Resistant *Staphylococcus Aureus* (MRSA) was low (7.1%), same result has been found in another study²³. However, the prevalence of MRSA in our study is remarkably low as compared to the study by Safia²⁴ which showed 100% prevalence of MRSA. Our study revealed that *Staphylococcus aureus* showed highest sensitivity (100%) to Vancomycin. Same result has been observed by Safia et al²⁴. Good sensitivity patterns were shown to amikacin (91%), Tetracyclines (87.5%), Ciprofloxacin (83.3%) and Gentamicin (78.5%). These findings are in agreement with the previous studies^{7,16,21}.

In our study moderate sensitivity of *Staphylococcus aureus* (57%) has been observed against Amoxicillin/clavulonic acid while a study done in Agha Khan University hospital indicated good sensitivity (69.76%) to it²¹. In present study, *S-aureus* showed low susceptibility patterns to 2nd and 3rd generation cephalosporines as compared to some studies^{21,25}. This could be due to the overuse of these drugs and the high prevalence of extended spectrum beta lactamases among these organisms. Our study indicated low susceptibility of *S-aureus* to Penicillin and

Ofloxacin. Same results have been observed in recent studies^{21,24,26}.

In one case, MRSA was detected which was sensitive to Vancomycin, Chloramphenicol and Linezolid, while it was resistant to Ciprofloxacin, Gentamicin, Clindamycin, Fusidicacid, Oxacillin and Penicillin. Same sensitivity pattern of MRSA have been observed in other studies carried out in different hospitals in Pakistan^{27,28,29}. In our study, *Pseudomonas aeruginosa* strains showed good sensitivity patterns to Imepenem (100%), Piperacillin-Tazobactam (87.5%), Amikacin (75%) and Ciprofloxacin (75%). These findings are in agreement with studies done by varsha¹⁵ and Shekih²¹. However, in our study low sensitivity patterns to 2nd and 3rd generation Cephalosporines have been seen. This result is in conformity with the studies in the literature^{21,24,26,30}.

The increased resistance against Cephalosporines might be the result of increased used of these antibiotics, as these are extensively used in the selected ward for prophylaxis as well as post operatively to prevent infections and as empirical treatment for infected cases. Literature also suggests that antibiotic use is proportional to the antibiotic resistance²⁴.

Sensitivity patterns of *Escherichia coli* in our study as compared to others were Amikacin (100% vs. 88.9%)²¹, Imipenem (100% vs. 78%)²⁴, Gentamicin (60% vs 51.8%)²¹ and Ciprofloxacin (80% vs. 70.4%).²¹ Good sensitivity patterns were observed to Cephalosporins i.e. Ceftriaxone (80%), Ceftazidime (60%) and Cefotaxime (60%). However, in contrary to our findings Safia and Ali³¹ have shown high resistance patterns to cephalosporins in their reports.

CONCLUSION

Infection control can be achieved by proper sterilization of the instruments, surfaces and theatres. Proper scrubbing techniques should be observed. Furthermore, Periodic surveillance is recommended to determine the susceptibility patterns of the common pathogens and tailoring the antimicrobials use accordingly.

REFERENCES

1. Prevention and treatment of Surgical Site Infection. National Collaborating Centre for women's and children health. Clinical guideline 2008: London RCOG Press.
2. Leape LL, Brennan TA, Laird N. The nature of adverse events in hospitalized patients. Results of the Harvard Medical Practice study. *N Engl Med* 1991; 324: 377-84.
3. Nathens AB, Cook CH, Machiedo G, Moore EE, Namias N, Nwariaku F. Defining the research

- agenda for surgical infection: A consensus of experts using the Delphi approach. *Surg Infect* 2006; 7: 101-10.
4. Ashby E, Haddad F S, O'Donnell E, Wilson AP. How will surgical site infection be measured to ensure high quality care for all? *Bone Joint Surg Br.* 2010; 92: 1294-99.
 5. Meo SA, Siddique S, Nawaz Q, Meo RA. Frequency and patient related risks for surgical site infection. *Pak Armed forces J.* 2011; 61: 259-61.
 6. Bayat A, MC Grouther DA, Ferguson MW. Skin Scarring. *British Med J* 2003; 326: 88-92.
 7. Sule AM, Thanni LOA, Sule Odu OA, Olusanya O. Bacterial pathogens associated with infected wounds in Ugan State University Teaching Hospital, Sagamu. *African J of clinical and experimental microbiology* 2002; 3: 13-16.
 8. Tiwari HK, Sen MR. Emergence of vancomycin resistant *Staphylococcus aureus* from a tertiary care hospital from northern part of India. *BMC Infect Dis.* 2006; 6: 156-59.
 9. Rahman J, Sultana N, Hassan M, Begum HA. Factors of Post operative wounds infection in abdominal surgeries of Obstetrics and Gynaecology department. *J Dhaka National Med. Coll. Hos.* 2011; 18: 39-42.
 10. Goswami NN, Trivedi HR, Goswami APP, Patel TK, Tripathi CB. Antibiotic Sensitivity Profile of bacterial pathogens in postoperative wound infections at a tertiary care hospital in Gujrat, India. *J Pharmacol Pharama cother.* 2011; 2: 158-64.
 11. Haley RW, Culver DH, White JW, Morgan WM, Emori TG, Munn VP, Hooton TM. The efficacy of infection surveillance and control programs in preventing nosocomial infection in US hospitals. *Am. J Epidemiol* 1985; 121: 182-205.
 12. Condon RE, Haley RW, Lee JT, Meakins JL. Does the infection control controls infections? *Arch. Surg.* 1988; 123: 250-56.
 13. Nisa M, Naz T, Irum A, Hassan L. Scope of surgical site infection in obstetrics and gynaecology. *J of Postgrad Med Inst* 2005; 19: 438-41.
 14. National Nosocomial Infections Surveillance (NNIS) System Report, data summary from 1992 to 2004. *Am J Infect Control.* 2004; 32: 470-85.
 15. Shahane V, Bhawal S, Lele U. Surgical Site Infections: A one year prospective study in a tertiary care center. *Int J of Health Sci* 2012; 6: 79-84.
 16. Anguzu JR, Olila D. Drug sensitivity patterns of bacterial isolates from septic post-operative wounds in a regional referral hospital in Uganda. *African Health Sci* 2007; 7: 148-154.
 17. Surucuoglu S, Ghazi H, Kurutepe S, Ozkutuk N, Ozbakkaloglu B. Bacteriology of surgical wound infections in a tertiary care hospital in Turkey. *East Afr Med J.* 2005; 82: 331-36.
 18. Lilani SP, Jangale N, Chowdhary A, Daver GB. Surgical site infection in clean and clean contaminated cases. *Indian J Med Microbiol.* 2005; 23: 249-52.
 19. Thanni LO, Osinupebi OA, Deji-Agboola M. Prevalence of bacterial pathogens in infected wounds in a tertiary hospital, 1995-2001: any change in trend? *J Natl Med Assoc.* 2003; 95: 1189-95.
 20. Giacometti A, Cicrioni O, Schimizzi AM, Delprete MS, Barchiesi F, D'Errico MM, et al. Epidemiology and Microbiology of Surgical Wound infections. *J Clin Microbiol.* 2000; 38: 918-22.
 21. Shaikh D, Zaidi A H, Shaikh K, Shaikh M, Naqvi B S. Post Surgical wound infections; A study on threats of emerging resistance. *Pak J of Pharmacology.* 2003; 20: 31-41.
 22. Naik G, Desphpande SR. A study on surgical site infections caused by *staphylococcus aureus* with a special search for Methicillin Resistant Isolates. *J of Clinical and Dig. Research.* 2011; 5: 502-8.
 23. Andhoga J, Macharia AG, Maikuma IR, Wanyonyi ZS, Ayumba BR. Aerobic Pathogenic bacteria in post operative wounds at Moi Teaching and Referral Hospital. *East Afr Med J.* 2002; 79: 640-44.
 24. Bibi S, Channa GA, Siddiqui TR, Ahmed W. Pattern of bacterial pathogens in postoperative wounds and their sensitivity patterns. *J of Surgery Pak.* 2012; 17: 164-67.
 25. Siegrist HH, Nepa MC, Jacquet A. Susceptibility to Levofloxacin of clinical isolates of bacteria from intensive care and hematological/oncology patients in Switzerland: a multicentre study. *J Antimicrob Chemother.* 1999; 43: 51-54.
 26. Khan MA, Ansari MN, Bano S. Postoperative wound infection. *Ind. J. Surg.* 1885; 48: 383-86.
 27. Kaleem F, Usman J, Hussain A, Omair M, Khalid A. Sensitivity pattern of methicillin resistant *staptgcococcus aureus* isolated from patients admitted in a tertiary care hospital of pallesten. *Iran J Microbiol* 2010; 2: 143-46.
 28. Malik N, butt T, Bari-A. Frequency and antimicrobial susceptibility pattern of methicillin resistant aureus. *J Coll. Physicinan surg. Pak* 2009; 19: 287-90.
 29. Taj Y, Abdullah FE, Kazmi SU. Current pattern of antibiotic resistance in *stephylococcus aureus* clinical isolated and the emergence of vancomycin resistance *J Coll Physicens Surg tall.* 2010; 20: 728-32.
 30. Rastogi V, Mishra PK, Bhatia S. emergency antimicrobial resistance in hospital a threat to public health. *Ind J of Community health.* 2012; 24: 260-63.
 31. Ali SA, Tahir SM, Memon AS, Shaikh NA. Pattern of pathogens and their sensitivity isolated from superficial surgical site infections in a tertiary care hospital. *J Ayub Med Coll.* 2009; 21: 80-82.